





Volume 1; Issue 1

Olive Characteristic Determination of Six Cultivar Candidates for Scratched Green Table Olive Production

Yasin Ozdemir^{1*}, Nesrin Aktepetangu², Seda Kayahn¹ and Ozge Keskinel³

¹Department of Food Technologies, Ataturk Central Horticultural Research Institute, Turkey

²Department of Fruit Breeding, Ataturk Central Horticultural Research Institute, Turkey

³Department of Food Engineering, Ankara University, Turkey

***Corresponding author:** Yasin Ozdemir, Department of Food Technologies, Ataturk Horticultural Central Research Institute, Yalova, Turkey, Tel no: 0226 814 25 20 – 1652; Fax: 0226 814 11 46; E-mail: yasin.ozdemir@tarim.gov.tr

Received Date: May 24, 2018; Published Date: June 21, 2018

Abstract

Variety was reported as most important factor for quality characteristics of table olive. So that researchers indented to develop new high quality olive cultivars. For this purpose Ataturk Central Horticultural Research Institute developed nearly 1500 olive genotypes by crossing Turkish, Spanish and Italian high quality table olive cultivars. 6 olive genotypes were selected as new cultivar candidates for green table olive production by researchers according to their high fruit yield, relatively resistance to disease, large homogeneous green ripening fruits. These 6 green olives were used as material in this study and Domat which extensively produced green table olive cultivar was used to comparison. This research includes harvesting the olives at green maturity, production of scratched green table olive and determination of the physical and chemical characteristics of produced table olives. Size and weight of fruit and seed, water, titratable acidity, pH, tissue hardness, color and oleuropein absorbance (K_{345}) value and salt analysis were applied to evaluate fruits

and determine their suitability for scratched green table olive production. Fruit weight and flesh to seed ratio of fresh olives were 5.18-7.65 g and 4.5-6.8 LT011 had highest fruit weight and LT001 had highest flesh to seed ratio. Result of this study showed that LT001 and LT011 had superior characteristics for scratched green table olive production and this research also provided required data forthe new cultivar registration procedure.

Keywords: Genotype; Oleuropein; Hardness; Domat

Introduction

Many of the traditional olive cultivars do not meet the requirements for these new olive growing regions [1] because olive industry has dramatically changed and new olive growing techniques and systems has been used in the last decades [2]. New cultivars prompted the development of olive breeding programs in the main

olive-producing countries based in intra-specific crossbreeding between cultivars of known merit aiming at combining the good qualities of the progenitors in some of the genotypes of the progenies [3,4]. The concept of quality in fruit products is wide, complex and dynamic. It implies a large number of attributes with different significance according to the interest and expectations of the different stakeholders of the chain, from producers to

Citation: Yasin Ozdemir, et al. Olive Characteristic Determination of Six Cultivar Candidates for Scratched Green Table Olive Production. J Nutri Diet Probiotics 2018, 1(1): 180003. consumers [5]. Fruit weight, flesh and seed ratio and texture hardness (especially after processing) were thought as important physical quality attribute and has great importance for table olive breeding programs [6,7]. So that this research is aimed to produce scratched green table olivesfrom 6 cultivar candidates and determinate their some physical and chemical characteristics.

Materials and Methods

In this study, olive of 6 cultivar candidates which were chosen by breeding researcher on the basis of their high productivity and resistance to diseases and low periodicity according to results of national cross breeding project (Obtaining New Olive Varieties by Crossing, 1990-2018) and Domat cultivar were evaluated. Those trees were planted at in 1.5 m x 3 m distance in olive genotype observation orchard of Ataturk Central Horticultural Research Institute (Yalova/Turkey) in 2001.Maturity index of olives were followed according [8] and olives were randomly handpicked in 2014-2015. Code of olives, their crossing combination and maturity index were given in Table1.

| Code | Crossing combination | Maturation index |
|-------|-----------------------------|---------------------|
| BE001 | Belle d'Espagne X Edinciksu | 1.2 |
| BE003 | Belle d'Espagne X Edinciksu | 1.2 |
| BE005 | Belle d'Espagne X Edinciksu | 0.9 |
| LE001 | Lucques X Edinciksu | 1.1 |
| LT001 | Lucques X Tavşanyüreği | 1.2 |
| LT011 | LucquesTavşanyüreği | 1.1 |

Table 1: Olives code numbers, their parents and their maturity index.

Method of table olive production

Olives were processed to starched green table olive according to method of, Turkish Food Codex Table Olive Communiqué Regulation [9]. Each olive was vertically scratched and kept in tap water. This water was changed daily with fresh tap water during 10 days. After those olives kept in 8% brinewhose pH was adjusted to 5 by addition of acetic acid to accelerate spontaneous fermentation and prevent growing of unwanted microorganism. Olives were fermented in brine at~ 16°C until pH fall to 4.4.

Physical and chemical analysis

Fruit weight, flesh to seed ratio, pH value, titratable acidity and sodium chloride content were determined according to official method of Turkish Table Olive Standard [10]. Water content of olive samples was determined in a conventional oven at 75±2°C [11]. Color values of olive skin were measured with a color meter (Konica Minolta, Japan). Texture hardness of olive was measured with fruit harness tester (W.O.W FRH-5, Japan). Absorbance value of oleuropein was determined by spectrophotometric method at 345 nm according to [12].

Statistical analysis

Randomized experimental design was used and analysis of variance was applied with the Duncan multiple comparison test of the means (p<0.01) to determine the presence of significant differences among the samples. Statistical analysis was performed by using the JMP v. 5.0 statistical package programs (SAS Institute, Cary, N.C., U.S.A.). Different letters indicate significant difference in same colon of tables.

Results and Discussions

Price of olive increase according to its fruit weight and flesh to seed ratio [13] so that, these values are required to be high for new olive cultivar. Flesh to seed ratio was required at least 5 for new table olive cultivar candidates by breeding researchers [14]. Fruit and seed size, fruit weight and flesh to seed ratio of olive were given in Table 2. In this study fruit weight and flesh to seed ratio was determined similar with result of [15] but higher than results of [16] and [17] for evaluated olive cultivars and cultivar candidates. In this research flesh to seed ratio of all table olives had higher than 5 except BE003 and LT011.

| Sample | Fruit length (cm) | Fruit width (cm) | Seed length (cm) | Seed width (cm) | Fruit weight (g) | Flesh to seed ratio |
|--------|-------------------|---------------------|---------------------|--------------------|------------------|------------------------|
| BE001 | 2.1 | 3.1 | 0.9 | 2.4ab | 6.2b | 5.0c |
| BE003 | 2.0 | 2.8 | 0.9 | 2.2c | 5.6c | 5.0c |
| BE005 | 1.9 | 3.1 | 0.9 | 2.6a | 5.6c | 4.0e |
| LE001 | 2.0 | 2.9 | 0.9 | 2.1c | 5.2d | 6.3b |
| LT001 | 2.2 | 2.9 | 0.9 | 2.0c | 6.2b | 6.8a |
| LT011 | 2.3 | 3.1 | 0.9 | 2.1c | 7.6a | 5.7bc |
| Domat | 2.2 | 2.9 | 0.9 | 2.2bc | 6.4b | 4.6d |

Table 2: Fruit and seed size, fruit weight and flesh to seed ratio of processed olive.

Glossy color and high flesh hardness of table olives was one of the required criteria [18,14]. According to surface color of olives distinguishes four elaboration types according to surface color: green, turning color, natural black and ripe olives [8]. So that in these research table olives was categorized in green olive. Color values, hardness, pH and titratable acidity of raw olive samples were given in Table 3. The hardness of aolive is its surface resistance to penetration of an indenter which is an important table olive quality criteria [14,19]. In this research hardness, pH and titratable acidity of olive samples were determined between 372-576 g, 4.31-4.48 and 0.14-0.23 % oleic acid. Titratable acidity contents of the olives were similar with results of [20,21].

| Sample | Color values | | | Hardness (g) | pН | Titratable acidity | |
|--------|--------------|-------|-------|--------------|------|--------------------|--|
| | L | а | b | naruness (g) | рп | (% oleic acid) | |
| BE001 | 39.19 | 0.56d | 14.00 | 556a | 4.42 | 0.23a | |
| BE003 | 36.84 | 0.98c | 16.01 | 506ab | 4.32 | 0.14bc | |
| BE005 | 39.56 | 1.05c | 18.23 | 497b | 4.37 | 0.21a | |
| LE001 | 38.29 | 0.55d | 16.02 | 372c | 4.31 | 0.15bc | |
| LT001 | 35.96 | 2.60a | 14.63 | 393c | 4.42 | 0.15bc | |
| LT011 | 38.62 | 1.37b | 14.58 | 461b | 4.48 | 0.14c | |
| Domat | 38.54 | 1.58b | 13.91 | 463b | 4.40 | 0.18b | |

Table 3: Color values, hardness, pH and titratable acidity of processed olives.

Water and salt content, oleuropein absorbance value of olive samples were given in Table 4. Oleuropein was important phenolic component and responsible from this bitter taste [22] so that debittering steps were used in table olive processes [23]. In this study oleuropein content of the olives was decreased by diffusion of oleuropein during daily changing water. Scratching also accelerated the water soluble content diffusion by crossing skin barrier of olives. Oleuropein absorbance value was used as an indicator of bitterness of olives [18,24]. Oleuropein absorbance value of the samples were in accordance with the previously published works of [23,25,26]. But oleuropein absorbance values were higher than the results of previous studies [22,27].

| Sample | Water (%) | Oleuropein absorbance value (K345) | Salt (%) |
|--------|-----------|------------------------------------|----------|
| BE001 | 69.83 | 0.33b | 2.27 |
| BE003 | 68.93 | 0.25d | 2.00 |
| BE005 | 70.62 | 0.31bc | 2.10 |
| LE001 | 66.26 | 0.28cd | 2.47 |
| LT001 | 69.49 | 0.36b | 2.17 |
| LT011 | 68.42 | 0.48a | 2.13 |
| DOMAT | 68.91 | 0.23d | 2.33 |

Table 4: Water and salt content and oleuropein absorbance value of processed olive samples.

Fermentation improved flavor and texture characteristics [27] and color as a consequence of the different pigment polymerization and phenol oxidation [28]. For production of spontaneous fermented table olives, the fruit should be at optimum ripeness according to selected production method to obtain excellent color and texture hardness after processing to attract consumer [27]. In fact, in this research olives were harvested between 0.9-1.2 maturation indexes and processed with spontaneous fermentation after debittering olives by daily changing water.

Conclusions

In this research 6 cultivar candidates from previously finished cross breeding project and 1 standard cultivar were studied. Seed width, fruit weight, flesh to seed ratio, color value of hardness, titratable acidity and oleuropein absorbance value evaluated criteria determined as statistical significant different characters of olives. Differences were explained by the genetic variation of these cultivar candidates because they cultivated and processed under same conditions. LT001 was also found to have highest fruit weight and flesh to seed ratio. As a result of this research, LT011 and LT001 were remarkable olives for scratched green table olive production and they can be advisable to breeding researcher for registration and certification.

References

- Rallo L, Caruso T, Díez C M, CampisiG (2016) Olive Growing in a Time of Change: From Empiricism to Genomics. In: Rugini E, et al. (Eds.), The Olive Tree Genome. (1st edn), Springer International Publishing, pp. 55-64.
- Rallo P, Morales-Sillero A, Brenes M, Jiménez MDR, Sánchez AH, et al. (2018) Elaboration of Table Olives: Assessment of New Olive Genotypes. European Journal of Lipid Science and Technology 120(6).
- 3. Sánchez de Medina V, Calderón-Santiago M, E Riachy M, Priego-Capote F, Luque de Castro MD (2014) High resolution mass spectrometry to evaluate the influence of cross-breeding segregating populations on the phenolic profile of virgin olive oils. J Sci Food Agric 94(15): 3100-3109.
- 4. Lorenzo León, Leonardo Velasco, Raúl de la Rosa (2015) Initial selection steps in olive breeding programs. Euphytica, 201(3):453-462.
- 5. Marios C Kyriacoua, Youssef Rouphael (2018) Towards a new definition of quality for fresh fruit and vegetables. Scientia Horticulturae 234: 463-469.
- 6. Rallo L (2014) Breeding oil and table olives for mechanical harvesting in Spain. HortTechnology 24(3): 295-300.
- Ozdemir Y, Akcay ME, Kurultay S (2011) Melezleme Islahı ile elde edilen iki amaçlı (sofralık ve yağlık) zeytin çesit adaylarına genel bir bakış. Bahçe 40(1): 29-36.
- 8. BOE (2001) Real Decreto. Reglamentacióntécnicosanitaria para la elaboración, circulación y venta de las aceitunas de mesa, 279(21): 42587-42594.
- Anonymous (2014) Table Olive Communiqué, Turkish Food Codex Regulation, Published in date and number: 23.08.2014-29097, Communiqué No: 2014/33.
- 10. TS 774 (1992) Turkish Standards Institute, Turkish Table Olive Standard. Ankara, Turkey.

- 11. Esti M, Cinquanta L, La Notte E (1998) Phenolic compounds in different olive varieties. J Agric Food Chem 46(1): 32-35.
- 12. Mastorakis M, Sotiroudis TG, Xenakis A, Miniadis-Meimaroglou S (2004) Spectrophotometric analysis of enzymic and non-enzymic oxidation of oleuropein. Chemistry and Physics of Lipids 130(1): 58-58.
- 13. Son L (2004) Effects of hand and chemical thinning on fruit size and quality of 'Priana'and 'Beliana'apricot (Prunus armeniaca) cultivars. New Zealand Journal of Crop and Horticultural Science 32(3): 331-335.
- 14. Varol N, Erten L, Turanlı T (2009) Zeytin. The TarımveKöyişleriBakanlığı Teşkilatlanma veDesteklemeGenelMüdürlüğü (52): 330.
- 15. Medina E, Gori C, Servili M, De Castro A, Romero C, et al. (2010) Main variables affecting the lactic acid fermentation of table olives. International Journal of Food Science & Technology 45(6): 1291- 1296.
- 16. Rallo L, Barranco D, De La Rosa R, León L (2008) 'Chiquitita'olive. HortScience 43(2): 529-531.
- 17. Arji I, Bahmanipour F (2014) Adaptation Ability of some Olive Cultuvars and Genotypes in Ilam Province. Seed and Plant Improvment Journal 30(4): 761-775.
- 18. Boskou D, Camposeo S, Clodoveo ML (2015) Table olives as sources of bioactive compounds. Olive and Olive Oil Bioactive Constituents, pp. 217-259.
- 19. Castro-Garcia S, Rosa UA, Gliever CJ, Smith D, Burns JK, et al. (2009) Video evaluation of table olive damage during harvest with a canopy shaker. HortTechnology 19(2): 260-266.
- 20. Piga A, Gambella F, Vacca V, Agabbio M (2001) Response of three Sardinian olive cultivars to Greekstyle processing. Ital J Food Sci 13(1): 29- 40.
- 21. Poiana Marco, Romeo FV (2006) Changes in chemical and microbiological parameters of some varieties of Sicily olives during natural fermentation. Grasas y aceites 57(4): 402-408.
- 22. Panagou EZ (2006) Greek dry-salted olives: monitoring the dry-salting process and subsequent physico-chemical and microbiological profile during storage under different packing conditions at 4 and 20 °C. LWT-Food SciTechnol 39(4): 323-330.

- 23. Ben Othman N, Roblain D, Thonart P, Hamdi M (2008) Tunisian table olive phenolic compounds and their antioxidant capacity. J Food Sci 73(4): 235-240.
- 24. Ozdemir Y, Guven E. Ozturk A (2014) Understanding the Characteristics of Oleuropein for Table Olive Processing. J Food Process Technol 5(5): 1-6.
- 25. Morello JR, Romero MP, Motilva MJ (2004) Effect of the maturation process of the olive fruit on the phenolic fraction of drupes and oils from Arbequina, Farga, and Morrut cultivars. J Agric Food Chem 52(19): 6002–6009.
- 26. Sahin I, Korukluoglu M, Gurbuz O (2002) Salamurasiyahzeytinişlemedeçesit, mayavelaktik starter kullanımıvebazıkatkılarınfermentasyonsüresi

veürünkalitesineetkilerininaraştırılması. Turkiye 7. Gida Kongresi 203–212.

- 27. Sánchez Gómez AH, García P, Rejano Navarro L (2006) Elaboration of table olives. Grasas Y Aceites 57(1): 86-94.
- 28. Romero C, Brenes M, Yousfi K, Garcia P, Garcia A, Garrido A (2004) Effect of cultivar and processing method on the contents of polyphenols in table olives. J Agric Food Chem 52(3): 479- 484.

Kumral A, Basoglu F, Sahin I (2009) Effect of the use of different lactic starters on the microbiological and physicochemical characteristics of naturally black table olives of Gemlik cultivar. Journal of food processing and preservation 33(5): 651-664.