





Volume 5 Issue 1

Light Spectra and 6-Benzylaminopurine in the *In Vitro* Cultivation of Epidendrum lilas

Luciana CNL*, Rocha SS, Calaes JG and Pimenta S

EMBRAPA Mandioca e Fruticultura, Brazil

***Corresponding author:** Luciana Cardoso Nogueira Londe, Empresa de Pesquisa Agropecuária de Minas Gerais - EPAMIG, Campo experimental do Gorutuba, Nova Porteirinha - MG, Brazil; Email: luciana@epamig.br

Received Date: February 01, 2021; Published Date: February 23, 2022

Abstract

In order to supply the growing market demand for orchids (Epidendrum Lilas), is usually recommended the micropropagation technique, mainly, for the production of quality seedlings in a short time and in large numbers. Since, the propagation of this species is slow through conventional means, factors such as the phytoregulators and the light spectrum that may interfere in the effectiveness of this technique. The objective of the work was to evaluate different concentrations of cytokinin and the use of different light spectra in the in vitro propagation of orchid Epidendrum lilas. Two factors were evaluated: concentrations of benzylaminopurine (BAP): (1.0; 1.5; 2.0; 2.5 and 3.0 mg L-1) and the light spectra: white, red, blue and green. After two months of establishment, the following characteristics were evaluated: plant height (H), number of leaves (NL), leaf length (LL) and shoots (S). The data obtained were submitted to analysis of variance and the means compared by the Tukey test (p <0.05). There was a significant difference only for the light spectra. The red spectrum was more efficient for the development of the aerial part and leaf length. While the blue spectrum promoted a greater number of leaves. Higher shooting rates were observed using the white spectrum. It is concluded that during the orchid growth phase the red and blue spectra are more efficient, and in the multiplication phase the white spectrum is recommended.

Keywords: Orchid; Micropropagation; Light; Cytokinin

Introduction

The commercial production of flowers and ornamental plants has increased over the years, in all regions of Brazil [1]. Highlighting the orchid trade, which generates a commercialized amount of approximately US \$ 20 billion per year [2]. Being highly appreciated by consumers, due to its exotic flowers of vibrant colors and floral longevity [3]. Despite the great demand in the market, the spread of orchid species is considered slow, requiring a long period to reach the reproductive stage, making the production of

new seedlings time-consuming [4]. In natural conditions, the multiplication of these plants occurs through the natural propagation of the seeds, which do not have a functional endosperm. So that, under natural conditions, they depend on mycorrhizal fungi for symbiotic germination, which are necessary until adulthood for their survival [5]. In addition, seeds have relatively low germination rates, around 5% [6]. In view of the importance of orchids in the economic sector and due to the difficulty of their natural propagation, it is necessary to seek techniques that help the rapid propagation of this species. The micropropagation technique being the

most used, since it allows the use of all the seeds produced and the regeneration of adult plants from them [7].

In in vitro growth, the nutrient media are supplemented with growth regulators [8]. Highlighting the cytokinins, responsible for cell division, stretching and differentiation [9]. Among the most used cytokinis in in vitro cultivation, stands out the benzylaminopurine (BAP) [10]. However, the addition of these growth regulators is a factor that increases the costs of in vitro seedling production. An alternative would be manipulation of the cultivation environment through spectral quality [11]. In which it has proven action on plant development [12]. Spectral quality is essential in in vitro propagation, since light is essential for photomorphogenesis in plants [13]. With the variation in the light spectrum, the in vitro growth of several species can be manipulated, in an alternative way to the addition of phytoregulators to the culture medium [14]. In view of these aspects, the objective of the work was to evaluate the development and multiplication of the E. Lilas orchid under different light spectra associated to BAP doses.

Materials and Methods

Location of the Experiment

The experiment was conducted at the Plant Biotechnology Laboratory of the Agricultural Research Company of Minas Gerais-EPAMIG, EPAMIG Norte-Gorutuba Experimental Field, Nova Porteirinha-MG, during the years 2018/2019. The geographical location is defined by the coordinates 15°148'263" south latitude and 43°17'650" west longitude, the average altitude is 526 m.

Seed Establishment in Vitro

Orchid plants were previously established in vitro using seeds from their fruits (capsules). The collected capsules underwent a disinfection process and then brought to a laminar flow cabinet in which they were opened and the seeds removed to perform in vitro sowing. These seeds were placed in bottles with solid medium MS [15], supplemented with 0.5 mg L⁻¹ of 6-benzylaminopurine (BAP), 30 g L⁻¹ of sucrose, 0.1 g L⁻¹ of inositol, with pH adjusted to 5.8 \pm 0.1. After sowing, the flasks containing the seeds remained in a growth room for 60 days, or until sprouting, under irradiance of 40 µmol m⁻² s⁻¹ provided by cold white light, with 16 hours of photoperiod at 25 \pm 2°C. After 60 days, sprouts were used for the experiment.

In Vitro Cultivation Conditions

The obtained sprouts were individualized and transferred to

300 mL flasks containing 50 mL of MS medium supplemented with the following doses of BAP: 0.5, 1.0, 1.5, 2.0, 2.4 and 3.0 mg L⁻¹. Then the explants were taken to the growth room and subjected to different light spectra: white spectrum (cold white fluorescent light), red, blue and green spectrum. To obtain the colored spectra, the white lamps were wrapped with colored films of regenerated cellulose (cellophane), in the red (~ 625-440nm), blue (~ 440-485nm) and green (\sim 500-565nm) colors. The material was kept in a growth room for 60 days at a temperature of $25 \pm 2^{\circ}C$ and 16 hours of photoperiod (1,800 LUX). At 30 and 60 days after in vitro establishment, plant height (cm), leaf number, leaf length (cm) and shoots were evaluated. The experiment was conducted in a completely randomized design in a 6x5 factorial scheme (Doses of BAP x Colored spectra), with five replications.

Statistical Analysis

The variables were submitted to the Shapiro-Wilk normality test and the Bartlett test, both p <0.05, to verify the normality of the data and homogeneity of the variances, respectively. All variables were transformed by the x + 1 root. After confirming the requirements for an analysis of variance, it was performed at a probability of 5% error. Significant differences were observed for the sources of variation involved in the experiment, the means of the variables were subjected to the Tukey test (p> 0.05) to detect the differences between treatments. All statistical analyzes were performed using the Genes software [16].

Results and Discussion

There was no significant interaction (p < 0.05) among the factors dose and light spectra for any of the characteristics evaluated at 30 and 60 days of cultivation. There was a significant difference only for single factor spectra for the following: plant height, number of leaves and shoots. For characteristic leaf length there was a significant difference only at 60 days of cultivation. There was no significant difference for BAP doses. This fact may be associated with the balance between endogenous and exogenous cytokinin, the endogenous being sufficient to supply the needs of the plants [17]. It can also be related to luminosity. According to Chee and Pool, [18] high light intensity leads to photoxidation causing degradation of cytokinins. The plants showed greater growth when submitted to the red spectrum, at 30 and 60 days of cultivation Table 1. Similar results were observed by Rocha, et al. [19] working with forage palm cv. Gigante (Opuntia ficus indica Mill.).

Advances in Agricultural Technology & Plant Sciences

Espectra	30 days			60 days			
	H (cm)	NL	S	H (cm)	NL	LL (cm)	S
Blue	4.86 ab	1.77 ab	1.31 b	5.57 ab	2.36 a	3.77 ab	1.78 b
White	4.48 b	1.99 a	1.65 a	5.26 ab	2.58 a	3.33 b	2.13 a
Red	5.43 a	1.79 ab	1.22 bc	5.94 a	2.07 b	4.1 a	1.45 c
Green	4.43 b	1.70 b	1.04 c	4.78 b	1.81 b	3.52b	1.11 d

Table 1: Height (H), number of leaves (NL), leaf length (LL) and shoots (S) means of orchid Epidendrum Lilas, at 30 and 60 days of in vitro cultivation under different light spectra.

Means followed by the same letter do not differ statistically by Tukey's test (p < 0.05) for each variable. Some studies have concluded that the red spectrum promotes the growth of the aerial part of plants [20]. This is because the process of light absorption by plants (photomorphogenic route) is similar to the mechanism of action of hormones [13]. Rosa, et al. [21] working with Dendrobium phalaenopsis obtained plants with a length of 3.33 cm using 3.0 mg L⁻¹ of BAP. Rodrigues, et al. [22] observed greater plant growth (2.36 cm) with 1.0 mg L⁻¹, BAP in Oncidium baueri. In this work, using only the red spectrum, the plants had a height of 5.43 and 5.94 cm at 30 and 60 days of cultivation, respectively. Based on the similarity of the responses, it appears that it is possible to manipulate the in vitro growth of orchids, in an alternative way to the addition of phytoregulators to the culture medium. Plants grown under white spectrum showed a higher number of leaves (1.99 cm), at 30 days of cultivation. And at 60 days, more leaves were obtained with the blue (2.36) and white (2.58) spectrum.

Similar results for these spectra were observed by Cunha, et al. [23], working with Mentha spicata and Santos, et al. [24] observed similar averages using 3.6 mg L⁻¹ of BAP in Epidendrum ibaguense, confirming the efficiency of the spectra. These results may be related to the influence of these spectra on in vitro photosynthesis. Since leaves irradiated with white light absorb more blue, red and green wavelengths, which are necessary for energy gains through photosynthesis, as well as for other physiological processes [25]. Blue light is also essential in the processes of synthesis of pigments, enzymes, development of chloroplasts, stomatal opening and closing and several other photomorphogenic processes [26]. That may have favored the orchid leaf formation process. Plants exposed to the red spectrum showed greater leaf length (4.10 cm), at 60 days of cultivation. The red light generally emits a spectrum near to the maximum absorption of chlorophylls and phytochromes, being important for the development of the photosynthetic apparatus and for the accumulation of starch [27].

Shooting induction in orchids occurs more intensely when grown in white spectrum, at 30 (1.65 shoots / explant) and 60 days (2.13 shoots / explant). Similar results were observed

by Ferrari, et al. [28] in Curcuma longa. Camargo, et al. [3] working with Oncidium baueri observed a close average (1.87 shoots / plant) using 2.0 mg L⁻¹ of BAP. In general, plants grown in white light, an environment that is similar to the natural environment, present thicker leaves with longer and juxtaposed palisade parenchyma cells [26]. This may have favored in vitro photosynthesis and, consequently a higher shooting rate. Plants grown under green spectrum showed less development. Studies with Arabidopsis have reported that the green light causes similar effects when the plants are shaded, that is, they lead to etiolation and cause differences in the architecture of the plant such as lengthening of petioles, reorientation of leaves and reduction of leaf area [30]. Different results were observed by Rocha, et al. [31] working with three banana cultivars (Musa sp.), observed higher shooting rates using green light. However, the influence of spectral quality in relation to plant growth and development may be directly linked to the species, the stage of plant development and other environmental characteristics [32]. Which may justify the different results verified in this work. Since there was no influence of the phytoregulator used, the luminosity was the determining factor for the in vitro development of the orchid. These results indicate new possibilities for the micro propagation of E. Lilas, and can be considered as a starting point for further research on protocols, involving the cultivation environment, in order to improve the quality of the seedlings produced.

Conclusion

Under the conditions in which this experiment was carried out, the following conclusions were reached:

- The light spectra influences in the in vitro development and multiplication of orchids.
- The red spectrum provides greater aerial part development and leaf length. Whereas, plants exposed under the blue and white spectra have a higher number of leaves.
- Cultivation in a growth room under a white spectrum induces higher shooting rates.
- For economic matter, it is recommended to use the red and blue spectra in the growth phase and the white spectrum in the Epidendrum Lilas multiplication phase.

References

- 1. Junqueira AH, Peetz MS (2014) O setor produtivo de flores e plantas ornamentais do Brasil, no período de 2008 a 2013: atualizações, balanços e perspectivas. Revista Brasileira de Horticultura Ornamental 20(2): 115-120.
- 2. (2016) International Trade Centre. Market Dynamics, Annual Report.
- Deka K, Sharma PB, Sarma B, Borthakur SK, Tanti B (2017) Preventing extinction and improving conservation status of Vanilla borneensis Rolfe-A rare, endemic and threatened orchid of Assam, India. Journal for Nature Conservation 37: 39-46.
- 4. Cardoso JC (2014) Publicação em cultivo in vitro de plantas: qualidade para o avanço científico e tecnológico. Horticultura Brasileira 32(4): 383-384.
- Chávez HK, Mosquera-Espinosa AT, Otero-Ospina JT (2015) Propagación in vitro de semillas de la orquídea Comparettia falcata Poepp. e Endl. (Orchidaceae) mediante técnicas simbióticas y asimbióticas. Acta Agronómica 64(2): 125-133.
- 6. Vudala SM, Padial A, Ribas L (2019) Micropropagation of Hadrolaelia grandis through transverse and longitudinal thin cell layer culture. South African Journal of Botany 121: 76-82.
- Silva CDS, Araújo LGD, Sousa KCI, Carvalho JCBD, Gonçalves LDA, et al. (2016) Cultivo in vitro de Epidendrum nocturnum (Orchidaceae) ocorrente no Cerrado da Região Centro-Oeste. Rodriguésia 67(4): 1083-1091.
- Sorgato JC, Rosa YBCJ, Soares JS, Lemes CSR, Sousa GGD (2015) Light in intermediate acclimatization of in vitro germinated seedlings of Dendrobium phalaenopsis Deang Suree. Ciência Rural 45(2): 231-237.
- 9. Murai N (2014) Review: Plant growth hormone cytokinins control the crop seed yield. American Journal of Plant Sciences 5(14): 2178-2187.
- Soares JS, Rosa YBCJ, Suzuki RM, Scalon SPQ, Rosa EJJ (2012) Germinação assimbiótica e desenvolvimento de Dendrobium nobile Lindl sob efeito de reguladores vegetais no tratamento pré- germinativo. Revista Brasileira de Plantas Medicinais 14(4): 617-623.
- 11. Dignart SL, Castro EMD, Pasqual M, Ferronato A, Braga FT, et al. (2009) Luz natural e concentrações de sacarose no cultivo in vitro de Cattleya walkeriana. Ciência e

Agrotecnologia 33(3): 780-787.

- Morini S, Muleo R (2003) Effects of light quality on micropropagation of woody species. In: Jain SM, et al. (Eds.), Micropropagation of woody trees and fruits. Dordrecht, Kluwer Academic Publishers, Pisa, Italy, p: 3-35.
- 13. Taiz L, Zeiger E, Moller IM, Murphy A (2017) Fisiologia e desenvolvimento vegetal. Artmed, Porto Alegre, Brasil, pp: 1- 888.
- 14. Braga FT, Pasqual M, Castro ED, Dignart SL, Biagiotti G, et al. (2009) Qualidade de luz no cultivo in vitro de Dendranthema grandiflorum cv. Rage: características morfofisiológicas. Ciência e Agrotecnologia 33(2): 502-508.
- 15. Murashige T, Skoog F (1962) A revised medium for rapid growth and bio assays with tobacco tissue cultures. Physiologia plantarum 15(3): 473-497.
- 16. Cruz CD (2016) Genes Software-extended and integrated with the R, Matlab and Selegen. Acta Scientiarum. Agronomy 38(4): 547-552.
- 17. Grattapaglia D, Machado MA (1998) Micropropagação. In: Torres AC, et al. (Eds.), Cultura de tecidos e transformação genética de plantas. Brasília, pp: 1-6.
- 18. Chee R, Pool RM (1989) Morphogenetic responses to propate trimming, spectral irradiance, and photoperiod of grapevine shoots recultured in vitro. Journal of the American Society for Horticultural Science 114(2): 350-354.
- 19. Rocha SS, Londe LCN, Calaes JG, Pereira JCG, Viana WS, et al. (2018) Effect of lighting spectrum and naphthaleneacetic acid (NAA) on in vitro development of cactus pear [Opuntia ficus-indica (L.) Mill]. Australian Journal of Crop Science 12(12): 1837-1843.
- Marks TR, Simpson SE (1999) Effect of irradiance on shoot development in vitro. Plant growth regulation 28(2): 133-142.
- 21. Rosa YBCJ, Reis CCA, Casemiro JCL, Soares JS, Sorgato JC, et al. (2015) Cultivation period, lighting conditions and BAP concentrations on in vitro induction shoots of Dendrobium phalaenopsis Deang Suree. Ornamental Horticulture 21(3): 323-330.
- 22. Rodrigues DB, Nadal MC, Camargo SS, Assis AM, Schuch MW, et al. (2016) Growth regulators and substrates for Oncidium baueri Lindl. micropropagation. Semina Ciências Agrárias 37(5): 2901-2910.

- Cunha SHB, Silva ST, Bertolucci SKV, Carvalho AA, Rocha TT, et al. (2019) Influência da qualidade de luz no crescimento e acúmulo de voláteis de Mentha spicata cultivada in vitro. Scientia Plena 15(9): 1-11.
- 24. Santos MRA, Ferreira MDGR, Marques MG (2016) BAP e AIB no cultivo in vitro de Epidendrum ibaguense KUNTH. Plant Cell Culture e Micropropagation 6(2): 90-98.
- 25. Araujo AG, Pasqual M, Rodrigues FA, Rodrigues JD, Castro EM, et al. (2009) Crescimento in vitro de Cattleya loddigesii Lindl. em diferentes espectros luminosos associados com ácido giberelico. Revista Ceres 56(5): 542-546.
- 26. Taiz L, Zeiger E (2004) Fisiologia do estresse. Fisiologia vegetal. Artmed, Califórnia, pp: 643.
- 27. Saebo A, Krekling T, Appelgren M (1995) Light quality affects photosynthesis and leaf anatomy of birch plantlets in vitro. Plant Cell, Tissue and Organ Culture 41: 177-185.
- 28. Ferrari MPS, Antoniazzi D, Nascimento AB, Franz

LF, Bezerra CS, et al. (2016) Espectros luminosos no desenvolvimento de plântulas de Curcuma longa cultivadas in vitro. Arquivos de Ciências Veterinárias e Zoologia da UNIPAR 19(4): 247-251.

- 29. Camargo SS, Rodrigues DB, Rodrigues CM, Assis AMD, Faria RTD, et al. (2015) Fitorreguladores e espectros de luz na micropropagação de Oncidium baueri Lindl. Ciência Rural 45(11): 2007-2012.
- 30. Zhang T, Folta KM (2012) Green light signaling and adaptive response. Plant Signal Behav 7(1): 75-78.
- 31. Rocha PSG, Oliveira RP, Scivitarro BM, Mosele SH (2017) Uso de LEDs na multiplicação in vitro de três cultivares de bananeira. Revista Colombiana de Ciências Hortícolas 11(2): 247-252.
- 32. Souza GS, Silva JS, Oliveira UC, Neto RBS, Santos AR (2014) Crescimento vegetativo e produção de óleo essencial de plantas de alecrim cultivadas sob telas coloridas. Bioscience Journal 30(1): 232-239.